REMARKS ON THE USE OF MARK-RECAPTURE METHODOLOGY IN ESTIMATING AVIAN POPULATION SIZE

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ABSTRACT.—Mark-recapture models are classified according to requisite assumptions about population closure. The resulting classes of models are briefly discussed and the experimental situations to which they apply are described. Model assumptions are presented, with emphasis being placed on those which are most likely to be violated in avian population studies. Comments are provided on experimental design, and previous ornithological studies in which the various models have been used are briefly reviewed. Special attention is devoted to the Jolly-Seber model which was developed for open populations and which perhaps has the greatest potential applicability to detailed, long-term population studies. A number of examples of the use of the Jolly-Seber model with avian mark-recapture data are presented in summary fashion.

The literature of field ornithology is much more extensive than that dealing with field studies of other terrestrial vertebrates. However, mark-recapture methods of estimating population size have seen only limited use in the ornithological literature, but are frequently employed in published studies of mammals, reptiles and amphibians. For example, reviews of small mammal population estimation are dominated by mark-recapture methodologies (e.g., see Smith et al. 1975), while such methods are barely mentioned in methodological reviews of avian population estimation (Kendeigh 1944, Berthold 1976, Shields 1979). We believe that the neglect of mark-recapture methods in avian studies results from the generally high visibility and audibility of birds and the relative ease with which they can be directly enumerated. This ease of enumeration has naturally and justifiably resulted in an emphasis by ornithologists on estimation methods that involve actual counts of birds (or nests), which are then expanded in various ways to estimate total population size or density. However, no single population estimation method is universally appropriate for avian studies, and there is still much debate about census methodology (see J. T. Emlen 1971, Berthold 1976, Shields 1979, this symposium).

We do not suggest that mark-recapture methods will generally provide a preferable alternative to direct observational methods of estimating avian population size. However, we do believe that mark-recapture studies can provide estimates that are useful in assessing the appropriateness of other estimation methods, and that they may provide the most reasonable means of estimating population size in some situations. In this paper we attempt to introduce the subject of mark-recapture methods and to briefly review their use in previous avian studies. We will be concerned only with the estimation of population size, and will thus omit discussion of models for estimating survival rate from band recoveries of dead birds (e.g., see North 1978 and reviews in Taylor 1966, Seber 1973, Brownie et al. 1978). Our discussion will be introductory and brief, and we urge the interested reader to consult the excellent general reviews of mark-recapture models and their associated literature provided by Cormack (1968, 1979) and Seber (1973).

Mark-recapture models can be usefully classified according to their requisite assumptions about population closure. We define a closed population as one which remains unchanged over the period of investigation and which is thus not influenced by mortality, recruitment, or migration (both emigration and immigration) during this time. An open population is one that does change over the period of investigation as a result of either mortality and emigration, recruitment and immigration, or both sets of factors.

CLOSED POPULATIONS

TWO-SAMPLE EXPERIMENT

The first attempt to estimate the size of a bird population using mark-recapture methods is generally attributed to Lincoln (1930; also see Winkler 1930), and the "Lincoln index" or "Petersen estimate" has been the most widely-used mark-recapture method for estimating animal population size. Although the assumption of population closure can be relaxed, the Lincoln index or estimate is generally modeled and discussed assuming a closed population. The Lincoln estimate is obtained from a two-sample experiment. A sample of \( n_1 \) birds is taken from a population of size \( N \). Individuals in the sample are marked, returned to the population, and allowed time to mix freely with the unmarked birds. A second sample of size \( n_2 \) is then taken, and the proportion marked in this sample \( m_2 \) out of \( n_2 \), where \( m_2 \) denotes the number of marked

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birds in the second sample) is assumed to estimate the proportion of marked birds in the population. The total population, \( N \), is then estimated based on this assumption:

\[
(n_2/n_1) = (n_1/N), \text{ so } \hat{N} = (n_1n_2/m_2).
\]

The above estimator is generally said to require the following assumptions (e.g., see Seber 1973:59): (1) the population is closed, (2) all animals have equal capture probabilities in the first sample, (3) marking does not affect subsequent catchability, (4) the second sample is a simple random sample, (5) animals do not lose their marks, and (6) all marked animals occurring in the second sample are reported. If \( n_1 \) and \( n_2 \) are fixed and if these six “ball-and-urn model” sampling assumptions hold, then the number of recaptures, \( m_2 \) (given \( n_1, n_2 \), and \( N \)), can be modeled with the hypergeometric distribution or (with either replacement sampling or large \( N \)) the binomial distribution. In both cases the maximum likelihood estimator of \( N \) is the same as the intuitive estimator (however this estimate is biased and can be modified as suggested by Chapman 1951 or Bailey 1951). If sample sizes are not fixed, then it seems reasonable to regard capture probabilities as fixed (either because of characteristics of the sampled animals or expenditure of fixed effort by the experimenter) and to model the numbers of individuals with each of the four possible capture histories (caught in period 1, caught in period 2, caught in both periods 1 and 2, not caught) as a multinomial random vector. The maximum likelihood estimator of \( N \) based on the multinomial model is again the same as the intuitive estimator, and its asymptotic variance is essentially the same as that obtained for the hypergeometric model (Cormack 1979). These two general types of models (fixed sample size hypergeometric and random sample size multinomial) also represent the most common approaches to modeling other mark-recapture experiments. Cormack (1979:220) briefly compares these two approaches and notes that “strict adherence to either sampling rule is seldom possible.” Although the biologist should be aware of these two common approaches (other approaches are also possible, such as the loglinear models of Fienberg 1972, Cormack 1979), the important practical result is that they have yielded essentially the same estimates for experiments to which they have both been applied.

The practical value of any estimate based on mark-recapture models will depend on how closely the field experiment and resulting data correspond to the assumptions of the chosen model and how precise (and accurate) the estimate is (e.g., as indicated by its estimated sampling variance), given that model assumptions are met. Both of these considerations, precision and assumption validity, are important when designing mark-recapture experiments and when interpreting their results. The problem of designing a two-sample experiment to estimate population size with specified levels of precision and accuracy is addressed thoroughly by Robson and Regier (1964; also see Seber 1973:64–70), and we recommend these references to ornithologists interested in planning such a study.

Assumptions generally stated for the Lincoln estimate were listed above, and a complete discussion of these assumptions and tests of their validity is presented by Seber (1973:70–104). Certain specific deviations from the assumption of population closure are permissible when using the Lincoln estimate. Of particular interest to ornithologists is the fact that members of a population may be subjected to mortality during the intersample period and, as long as the average mortality rates of marked and unmarked birds are the same, the Lincoln estimate, \( \hat{N} \), is still a reasonable estimate of initial (at the time of the first sample) population size (see Seber 1970b). The presence of both recruitment and mortality results in overestimation of both initial and final population size. Probably the best practical means of insuring that violations of closure are negligible involve using a short time period between samples and choosing an appropriate time of the year for sampling. Naturally, short intervals between samples provide less opportunity for mortality, recruitment, and migration than longer intervals. Similarly, experiments should be conducted at a time of the year when migration and recruitment of young birds to the population are not occurring.

Assumptions 2, 3 and 4 involve catchability, which is an important consideration in any mark-recapture study. Assumptions 2 (all animals having equal capture probabilities in the first sample) and 4 (the second sample is a simple random sample) provide good examples of the difficulties involved in applying ball-and-urn statistical models to biological populations. Among avian studies, there are numerous examples in which all individuals in a population are not equally catchable. Differences in catchability or the probability of being sampled are sometimes associated with classes or subgroups of birds. For example, evidence of sex-specific differences in trappability has been provided for numerous waterfowl species by Petrides (1944) and Bellrose et al. (1961). Sex-specific differences in the probabilities of both capturing and resighting Northern Fulmars (Fulmarus glacialis) have been noted by Dunnett and Ollason (1978). Band recoveries by hunters are sometimes used in...
two-sample mark-recapture experiments (e.g., Lincoln 1930, Bergerud and Mercer 1966, Geis 1972) and recovery rates of many species of waterfowl are known to differ sex-specifically (e.g., see Bellrose et al. 1961). Many bird species exhibit age-specific differences in the probability of being taken in both trap and hunting samples (e.g., see Farner 1949, Bellrose et al. 1961, Kautz 1977, Sulzbach and Cooke 1978). Catchability can also be associated with breeding status in some species (e.g., see Orians 1958, Robel 1969, Sulzbach and Cooke 1978). In any case, when capture probabilities differ among identifiable subgroups of animals, then it is appropriate to treat each subgroup separately when estimating population size.

Catchability assumptions can also be violated by individual variation in capture probability that is not associated with identifiable subgroups (see assumptions 2 and 4). It is very difficult, if not impossible, to draw inferences about individual variation in capture probability using data from two-sample experiments, but recapture data on individuals from experiments involving a number of sample periods (K-sample experiments) have been used to examine catchability in avian studies. Evidence of individual variation in capture and/or resighting probabilities has been provided for several bird species (e.g., Borror 1948; Orians 1958; Young 1958, 1961; Taylor 1966; Carothers 1979). When inherent variation in capture probability exists, then the first sample will consist of a high proportion of the more trappable animals. Therefore, the average catchability of the marked animals will be higher than that of the unmarked animals, and marked animals will tend to be overrepresented in the second sample, resulting in biased estimates, \( \hat{N} \). However, it is conceivable that an inherent high probability of being sampled by one method would not necessarily insure a high sampling probability for another method. As Seber (1973) has pointed out, two selective samples can provide an unbiased estimate, \( \hat{N} \), if the methods of selection are independent. In practice, this argument suggests that the use of completely different capture techniques for the first and second samples can provide some protection against bias resulting from variation in catchability. With birds, it is sometimes possible to use traps to obtain the first sample and to apply highly visible markings (e.g., colored leg or neck bands, patagial tags, body dyes, backtags; see Cottam 1956, Taber and Cowan 1969, Marion and Shamis 1977, for reviews of avian marking techniques). The second sample is then taken by observing birds and tallying resightings of marked individuals as well as sightings of unmarked birds. This approach has been used successfully with Mallards (Anas platyrhynchos), Blue-winged Teal (Anas discors) and Wood Ducks (Aix sponsa) (Cowardin and Higgins 1967) and with Willow Ptarmigan (Lagopus lagopus) (Bergerud and Mercer 1966). When dealing with hunted species it is possible to obtain the first sample via trapping and to then use band recoveries returned by hunters as the second sample (e.g., see Lincoln 1930, Bergerud and Mercer 1966, Moisan et al. 1967, Goudy et al. 1970, Geis et al. 1971, Robel et al. 1972, Whitcomb 1974).

Effects of trapping and handling on future capture probability can also result in model assumption violations (assumptions 3 and 4), and such effects have been noted in birds. Sometimes such effects result from a behavioral response to trapping. Evidence of “trap-happy” birds that tend to return frequently to baited traps (presumably to take advantage of easily obtained food) has been provided by Borror (1948), Taylor (1966), and Murton et al. (1972). Trap and net aversion or “shyness” have been noted in a number of species (see Borror 1948, Young 1958, Stamm et al. 1960, Taylor 1966). Presumably, this involves an adverse reaction to being trapped and handled that results in unusual wariness of the bird or that causes the bird to avoid the immediate vicinity of the traps (e.g., see Owen and Morgan 1975). In addition it is possible that birds could be injured during the capture and handling processes, and that this might result in lower survival probabilities for marked birds. Aversive behavioral responses and injuries can be minimized through the use of proper capture and handling techniques (see reviews in Taber and Cowan 1969, U.S. Fish and Wildlife Service and Canadian Wildlife Service 1977). Apparent cases of trap-happy and trap-shy birds can also be produced by certain methods of trap placement. For example, Swinebroad (1964) mist-netted a Wood Thrush (Hylocichla mustelina) population and obtained evidence of unequal capture probabilities which he attributed to net placement. Apparently, some birds in the study area were exposed to nets within their “maximum activity areas” while other birds were only exposed to nets on the fringes of their ranges. The marks or tags that are applied to birds can also affect survival and the probability of future recapture. For example, the accumulation of ice on neckbands and nasal saddles of geese during severe winter weather is a potential problem, but is not believed to affect survival to a great extent (e.g., see Greenwood and Bair 1974, Craven 1979). Neckbands have been reported to contribute to starvation in Snow Geese (Anser caerulescens) (Ankney 1975). Nasal discs and
saddles have been suspected of increasing mortality of diving ducks (see discussion in Bartonek and Dane 1964), and patagial tags have been suspected of increasing susceptibility of birds to predation and altering behavior patterns of tagged birds (Anderson 1963). In addition to causing mortality differences between marked and unmarked birds, some marking methods can produce unequal catchabilities of the two groups at the time of the second sample. For example, higher resighting probabilities have been suspected for patagial-tagged (Cowardin and Higgins 1967) and backtagged and dyed birds (Bergerud and Mercer 1966), than for unmarked birds.

Assumption 5 is that animals do not lose their marks. In most two-sample experiments the period between samples will be sufficiently short that loss of marks commonly applied to birds should not be a problem. Retention times for several types of short-term avian markers (e.g., dyes, backtags) are presented by Taber and Cowan (1969) and Marion and Shamis (1977). Avian leg bands are generally considered to be good long-term markers but can be lost because of excessive wear or removal by the banded bird. The problem of band loss is thought to be important in long-lived birds and has been reviewed by Farner (1955), Ludwig (1967), and Marion and Shamis (1977). Leg band “survivorship curves” were developed by Ludwig (1967) for gulls and terns based on wear and weight loss data on known age bands from recaptured birds. A method for estimating the probability of tag loss from double-tagged animals (i.e., animals to which two tags are applied) is provided by Seber (1973:94–96). A practical means of reducing band loss problems is to replace worn bands on captured birds.

Assumption 6 is that all marked animals occurring in the second sample are reported, and this assumption is generally relevant only to experiments in which the second sample is based on band recoveries made by the general public (e.g., hunting recoveries). In cases where all recovered bands are not reported, some workers have estimated the total number of recovered bands for use in Lincoln index estimates (e.g., see Moisan et al. 1967, Goudy et al. 1970, Geis et al. 1971). This procedure requires an estimate of the “reporting rate” or the proportion of recovered bands that is reported. Reporting rate has been estimated using either additional information on the number of recovered bands obtained from hunter questionnaire surveys (Geis and Atwood 1961, Martinson 1966, Martinson and McCann 1966), or “reward band” studies in which some bands are marked with a message that a reward is offered for their return and are assumed to have a reporting rate of 1.0 (Bellrose 1955, Tomlinson 1968, Henny and Burnham 1976).

There are a number of reports in the avian literature of the use of the Lincoln index to estimate population size, and here we will only list some representative examples. Borror (1948), Boyd (1956), Stamm et al. (1960), Nunneley (1964), Taylor (1966) and Robel et al. (1972) have computed Lincoln index estimates from trapping and netting samples. All of these authors exhibited appropriate concern for at least some of the assumptions of the method, and none of them rejected the method as being completely inappropriate for their experimental situations. In an interesting comparison of methodologies Stamm et al. (1960) obtained general agreement between Lincoln index estimates and spot-mapping (Williams 1936) estimates for a number of eastern deciduous forest bird species. As noted earlier, Lincoln index estimates have also been computed from resightings of dyed or tagged birds (e.g. Bergerud and Mercer 1966, Cowardin and Higgins 1967, Readshaw 1968). Bergerud and Mercer (1966) compared estimates from several methods with “direct counts” believed to be accurate within 5% of the true population. The Lincoln index based on resightings provided estimates that agreed well with the direct counts for three of four years, while other population estimation methods proved less reliable. Cowardin and Higgins (1967) thoroughly examined Lincoln index assumptions and concluded that emigration of marked birds and increased visibility of marked birds may have resulted in biased estimates. Nevertheless, they concluded that Lincoln index estimates of population size were more realistic than total counts of birds.

Hewitt (1963, 1967) suggested an interesting technique for obtaining Lincoln index estimates for Red-winged Blackbirds (Agelaius phoeniceus) which does not require the actual capture of birds. Territorial males seen along roads are “marked” using tape-recorded descriptions of their exact sighting locations. The road is traveled again, and the presence of a territorial male in a location which was occupied at the time of the first sample is considered as a resighting. Consideration of requisite assumptions and comparison of estimates with estimates obtained using other methods led Francis (1973) and Albers (1976) to conclude that Hewitt’s methodology will generally underestimate population size. Lincoln index estimates based on hunting recoveries of banded birds have been used frequently (e.g., Lincoln 1930, Crissey 1963, Bergerud and Mercer 1966, Moisan et al. 1967, Gou-
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and Mercer (1966) have reported that such es-
timates for Mallards and Willow Ptarmigan, re-
spectively, agreed well with other estimates of
capture probabilities to vary from one sampling
period to another, but assumes that
within a sampling period, all individuals have
the same capture probability. The general max-
imum likelihood estimator for \( N \) under \( M_n \) cannot be written in
closed form, but must be computed numerically.
As noted earlier, there is a great deal of evidence
of variation in capture probability among
subgroups and individuals of various bird
species. In addition, it is reasonable to expect
capture probabilities to vary from one sampling
period to another as a result of such factors as
weather conditions, changes in sampling effort,
etc.

Model \( M_0 \) of Otis et al. (1978; also see Schna-
bel 1938, Darroch 19) has historically been the
most commonly used model for \( K \)-sample
closed population mark-recapture experiments
(see reviews in Cormack 1968, Seber 1973). \( M_0 \)
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able in closed form. While \( M_0 \) is somewhat more
realistic than \( M_n \), its assumption of equal cap-

K-SAMPLE EXPERIMENT

Here we discuss experiments involving two or
more trapping or sampling periods. The two-
sample Lincoln index experiment is simply a
special case of this type of experiment, and we
treated it separately only because of its frequent
use relative to other mark-recapture experi-
ments. Otis et al. (1978) have presented a unified
treatment of population estimation from \( K \)-sam-
ple experiments with closed populations. We

The sampling scheme is very similar to that of
the two-sample experiment. Animals are cap-
tured during an initial sampling period, marked, and
returned to the population. A second sample
is then taken (e.g., on the following day) and
recaptures of marked animals are noted. New
captures are also given marks and all animals
are returned to the population. The procedure
is repeated for \( K \) sampling periods. Perhaps the
greatest operational difference between the \( K \)-
sample and the two-sample experiments is that
animals generally must be given individual
marks (e.g., serially numbered leg bands) in the
\( K \)-sample experiment. The models used to
describe recapture data from \( K \)-sample experi-
ments generally require complete capture his-
tories of individual animals for estimating
population size. The probability distribution for
the set of possible capture histories is then ex-
pressed using a multinomial model (hypergeo-
metric approaches have also been used in some
cases) treating population size and capture
probabilities as parameters.

Assumptions required by the models reviewed
here are that (1) the population is closed, (2)
animals do not lose their marks during the ex-
periment, and (3) all marks are correctly noted
and recorded at each trapping occasion. In ad-
dition to these three assumptions, each of the
models discussed by Otis et al. (1978) embodies
a different set of assumptions about sources of
variation in catchability or probability of cap-
ture. As indicated for the two-sample experi-
ment, proper choice of the time (season of the
year) and duration of the experiment are two of
the most important practical means of approach-
ing closure (assumption 1). Regardless of the
precautions taken in designing the experiment,
however, it is desirable to test this assumption
after the experiment is completed in order to
insure that closed population models are appro-
priate. Closure tests based on specific closed
population models are provided by Pollock et al.
(1974) and Otis et al. (1978:66–67). Care should
be used in interpretation of the results of the
Otis et al. (1978) closure test, however, because
it is sensitive to various sorts of variability (e.g.,
over time) in capture probabilities as well as to
non-closure. It is thus prone to false rejection of
the closure assumption. The assumption (2) that
animals retain marks should be easily met in
most closed population experiments because of
their short duration relative to retention times
of most types of marks. The correct recording
of marks (assumption 3) will be a natural con-
sequence of careful field work and can be in-
sured by the use of well-designed field data
data sheets and appropriate editing procedures for
keypunched or summarized data.

The simplest and least realistic model de-
scribed by Otis et al. (1978) is \( M_0 \). In addition
to the three assumptions listed previously, \( M_0 \)
assumes that all individuals in the population
have identical capture probabilities for each of
the \( K \) sampling periods. \( M_0 \) thus includes only
two parameters; population size, \( N \), and capture
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ture probability within a sampling period will still be inappropriate for the many bird species that exhibit individual variation in catchability and variation associated with nonidentifiable subgroups.

Model \( M_h \) (Otis et al. 1978) corresponds to the situation in which the initial capture of an individual affects its probability of capture on subsequent sampling periods. As noted earlier, trap and net responses have been well-documented in birds, and this model may thus be realistic for some bird species. The model contains three parameters: population size, capture probability for initial captures (i.e., for unmarked animals), and capture probability for subsequent captures (i.e., for marked animals). The capture probabilities for marked and unmarked animals are assumed constant for all time periods. A numerical procedure is again needed to compute the estimate, \( \hat{N} \).

Model \( M_h \) (Burnham and Overton 1978, Otis et al. 1978) corresponds to the situation in which each member of a population is characterized by a distinct capture probability. These individual capture probabilities are assumed to remain the same over all sampling periods. In the development of the estimation procedure, Burnham and Overton (1978) recommend treating the set of individual capture probabilities as a random sample of size \( N \) from some probability distribution. They then develop an estimator, \( \hat{N} \), using an extension of the jackknife method of bias reduction.

Models \( M_l, M_b, \) and \( M_h \) of Otis et al. (1978) represent efforts to model specific sources of variation in capture probability. However, we might reasonably expect more than one of these sources of variation to be important in a given experiment. For this reason, Otis et al. (1978) developed a set of models corresponding to the various combinations of these sources of variation in capture probability. An estimator for \( N \) can be obtained for one of these models, \( M_{bh} \), which assumes that each member of a population has a specific probability of capture prior to its initial capture and another specific probability of capture after it has been marked. No estimators for \( N \) are currently available for the other combination models (\( M_{hb}, \ M_{bh}, \ M_{bhh} \)).

Assumptions 1–3 for all of the models have been briefly discussed. In addition, the assumptions distinguishing between the models concern the type of variability present in the capture probabilities. Otis et al. (1978) suggest seven tests for evaluating these assumptions. Some tests compare two competing models or hypotheses (one of which is more general than the other), while the others assess the goodness-of-fit of a particular model to the data. Otis et al. (1978) have also developed a discriminant classification function based on data simulated from each of the eight models. This classification function can be used to provide an objective selection of the appropriate model (and thus the appropriate catchability assumptions) for a given data set. We note that other tests dealing with variation in catchability are available (e.g., see Young 1958, 1961, as well as later discussion of open population models). Suggestions about the practical aspects of designing \( K \)-sample experiments for closed populations are provided by Otis et al. (1978:74–80).

We are aware of only one published mark-recapture experiment with birds in which one of the \( K \)-sample closed population models described above has been used (see Hewitt 1967). Most of these models are relatively new and would not be expected to have received much use, but others (e.g., \( M_l \)) have been available for some time. Otis et al. (1978) have developed a comprehensive computer program (see White et al. 1978) which computes estimates for their models, computes test statistics, and uses the classification function to select the appropriate model. We suspect that the availability of this program will result in use of the Otis et al. (1978) models in future mark-recapture studies of birds. T. J. Dwyer (pers. comm.) is currently using the program in conjunction with his mark-recapture study of Woodcock (Philohela minor) at the Moosehorn National Wildlife Refuge, Maine. Preliminary results suggest that model \( M_l \) may be appropriate for some of Dwyer’s within-season data sets.

Under certain assumptions, models for \( K \)-capture experiments on closed populations can be based on the observed frequencies of capture (i.e., on the numbers of animals captured \( 1, 2, \ldots, K \) times). For example, if probability of capture remains constant over time (the assumption of \( M_n \), then the probability that an animal is captured \( x \) times is given by the binomial distribution (e.g., see Eberhardt 1969, Seber 1973:169–170). Since the number of animals in the population that are never caught \( (x = 0) \) is unknown, the observed frequency of recaptures follows the zero-truncated binomial distribution. Under this assumption the number of unobserved animals can be estimated (Seber 1973:169–170) and the fit of the data to the model can be evaluated using a \( \chi^2 \) goodness-of-fit test. If capture probability is constant and small, then the zero-truncated Poisson distribution provides a reasonable approximation to the binomial model for capture frequencies. Using the Poisson model, the zero class can again be estimated (see Eberhardt 1969, Seber 1973:170) and the fit of the model tested. If probability of capture is
not the same for each individual, then certain assumptions can lead to different truncated models of recapture frequency. For example, different hypotheses about the distribution of catchability can lead to geometric and negative binomial models (see reviews in Eberhardt 1969, Seber 1973).

Although, as Seber (1980) has suggested, model $M_b$ of Burnham and Overton (1978) will probably replace other capture frequency approaches, these methods have seen limited use in avian studies. Bergerud and Mercer (1966) used a truncated Poisson model to estimate ptarmigan population size. Eberhardt (1969) examined the fit of the truncated Poisson and geometric distributions to published trapping and sighting data of several bird species. Condor (Gymnogyps californianus) sighting data and Bobwhite Quail (Colinus virginianus) capture data fit both Poisson and geometric models (Eberhardt 1969). North (1978) examined the fit of Manx Shearwater (Puffinus puffinus) recapture data within years to Poisson, geometric and negative binomial models. North (1978) also used a “double Poisson” model to deal with trap response and a “heterogeneous model” to handle two classes of birds that differ in their response to initial capture. The geometric and heterogeneous models both produced reasonable estimates. Both Eberhardt (1969) and North (1978) suggested that in cases where two models appear to fit the data well, selection of the model to use must be based on a detailed consideration of the biological assumptions involved. However, there is not always a biological basis for choosing among competing models. Cormack (1979:228) points out that several possible models may fit capture frequency data well but yield vastly different estimates of population size. For example, Dwyer (pers. comm.) found that both geometric and Poisson models fit his Woodcock capture frequencies well in many cases, but that the two models produced very different estimates of population size. Because of this problem and because the variety of models in Otis et al. (1978) correspond to most of the different biological situations that have been used to derive other capture frequency models, we suspect that the Otis et al. models will generally be preferred in future $K$-sample closed population experiments.

OPEN POPULATIONS

Completely Open Populations

In this section we examine $K$-sample experiments that are conducted in the same general manner as those already discussed, but where population gains and/or losses are allowed to occur between sampling periods through birth/immigration and death/emigration. Early models for these open populations incorporated the view that population changes were deterministic, and that randomness was associated only with the sampling process (see review of these models in Cormack 1968). However, it was recognized that the change in population size and composition from one time period to another could be more realistically viewed as a stochastic process itself (e.g., see Robson 1969). This problem was partially solved by Darroch (1959), when he developed models for partially open populations; i.e., ones in which either gains or losses in population size, but not both, are allowed to occur. The solution for the completely open population stochastic model was presented simultaneously by Jolly (1965) and Seber (1965) (denoted the Jolly-Seber model), and their model has now virtually replaced the deterministic models in experiments on open populations.

Assumptions that are generally listed for the completely open model are: (1) every animal in the population has the same probability, $p_i$, of being caught in sample $i$, given that it is alive and in the population during sampling period $i$, (2) every marked animal in the population has the same probability, $p_i$, of surviving from sampling periods $i$ to $i + 1$, given that it is alive and in the population immediately after the time of release in sample $i$, (3) every animal caught in sample $i$ has the same probability, $v_i$, of being returned to the population, (4) marked animals do not lose their marks, (5) all marks are reported and correctly recorded on recovery, (6) all samples are instantaneous (i.e., sampling time is negligible), and (7) losses to the population from emigration and death are permanent. Assumptions 1–6 are listed in Seber (1973:196) and assumption 7 is noted by Robson (1969:126) and Seber (1973:199).

Both Jolly (1965) and Seber (1965) used multinomial approaches to modeling. In Jolly’s (1965) general formulation, $p_i$, $v_i$, and $v_i$ are considered as unknown parameters, and $N_i$, $M_i$ (the number of marked individuals in the population at sampling period $i$), $B_i$ (the number of new individuals entering the population between sampling periods $i$ and $i + 1$) and $p_0(M_i/N_i)$, or the proportion of individuals in the population at time $i$ which is marked) are treated as unknown random variables.

The estimate for population size under the Jolly-Seber model is:

$$N_i = M_i n_i/m_i \quad (i = 2, 3, \ldots, K - 1)$$

where $n_i$ is the number of animals caught at time $i$ and $m_i$ is the number of marked animals caught at time $i$. This is an intuitive Lincoln-type esti-
mals in the entire population \((M_i/N_i)\) is simply in the sample \((m_i/n_i)\). However, while \(n_i\) and \(m_i\) are observable random variables; \(M_i\) must be estimated. As pointed out by Jolly (1965; also see Cormack 1968, 1972) the estimator for \(M_i\) is also intuitively appealing:

\[
\hat{M}_i = \frac{R_i z_i}{r_i} + m_i \quad (i = 2, 3, \ldots, K - 1)
\]

where \(R_i\) is the number of marked animals released into the population after sampling period \(i\), \(r_i\) is the number of marked animals released after sampling period \(i\) and caught subsequently, and \(z_i\) is the number of animals caught before sample \(i\), but not in sample \(i\), and caught again after sample \(i\). The marked individuals in the population just after sampling period \(i\) consist of two groups: the animals captured during period \(i\) \((R_i)\), and the number not captured during period \(i\) \((M_i - m_i)\). Of the former group, \(r_i\) are subsequently recaptured, and \(z_i\) from the latter group are subsequently recaptured. The two ratios, \(r_i/R_i\) and \(z_i/(M_i - m_i)\), should be approximately equal and the solution of the equality for \(M_i\) yields \(\hat{M}_i\).

We are aware of no published guidelines for planning a mark-recapture experiment to be analyzed with the Jolly-Seber model. However, the form of the asymptotic variances, as well as the simulation studies of Manly (1970, 1971a), Gilbert (1973), Bishop and Sheppard (1973) and Kreger (1973), demonstrate the importance of high capture probabilities, \(p_i\), to obtaining accurate estimates (having low or negligible bias) of population size with low estimated variances. Gilbert's (1973) simulations also demonstrate that substantial reductions in bias of \(\hat{N}_i\) can result from increasing the length of the experiment (i.e., the number of sampling periods). The asymptotic variance estimators of Jolly (1965) are known to produce confidence intervals with poor coverage in some cases (Manly 1971a). Specifically, in small sample size situations there is a high positive correlation between \(N\) and its estimated variance, causing underestimates of population size to appear to be more precise than they really are (Manly 1971a). This problem has led to a search for alternate methods of estimating variances for this model (Manly 1977a).

Nevertheless, estimated coefficients of variation (standard error of estimate/estimate) of population size do provide some indication of how precise and reliable the estimates are (Kreger 1973). Carothers (1973) points out that an estimate with large "small-sample" bias can normally be recognized as unreliable by its large estimated variance.

We prepared Figures 1 and 2 in an effort to provide some indication of the sampling effort required to achieve various levels of precision of \(\hat{N}_i\). Figure 1 assumes a constant intersample survival rate of \(\phi_i = 0.50\) while Figure 2 assumes \(\phi_i = 0.75\). It was assumed that there were no accidental deaths during banding and handling. In both Figures, the \(N_i\) were assumed constant at values of 100, 200, 500 and 1000, and \(B_i = (1 - \phi_i)N_i\) in order to insure a stable population. All experiments were assumed to include six sampling periods. The solid lines in the Figures represent approximations to the expected value of the estimated coefficient of variation of \(\hat{N}_i\), \(\text{E}[\text{CV}(\hat{N}_i)]\). These were obtained by approximating the expected values of \(\hat{N}_i\) and the standard error of \(\hat{N}_i\), \(\text{SE}(\hat{N}_i)\). This was done by using the expected values, \(E(R_i)\), \(E(n_i)\), \(E(m_i)\), \(E(z_i)\), and \(E(r_i)\), given the \(N_i\), \(\phi_i\), \(B_i\) and \(p_i\), in conjunction with the equation for \(\hat{N}_i\) presented earlier and the equation for \(\text{var}(\hat{N}_i)\) presented by Jolly (1965). The approximation for \(\text{E}[\text{CV}(\hat{N}_i)]\) was then obtained as \(\text{E}[\text{SE}(\hat{N}_i)]/\text{E}[\hat{N}_i]\). The plotted points in Figures 1 and 2 each represent the mean value of \(\text{CV}(\hat{N}_i)\) computed from 500 simulated data sets. Simulated data sets were constructed by treating capture and survival of each individual for each sampling period as independent Bernoulli trials using pseudorandom numbers. If the selected values of \(\phi\) and \(N\) are thought to be reasonable guesses for certain populations, then Figures 1 and 2 may be of use in planning experiments.

Figures 1 and 2 illustrate the importance of high capture probabilities (and corresponding large sample sizes) to obtaining precise estimates of population size. Cormack (1979) has noted that the generality of the Jolly-Seber model, with its separate parameters for each survival and capture probability, may sometimes limit its practical utility. However, Jolly (1979, in prep.) has recently developed models (and appropriate model testing procedures) in which survival probability or capture probability or both are constant over time (i.e., \(\phi_i = \phi\), \(p_i = p\)). We suspect that these reduced-parameter models will be of considerable practical value.

Assumption 1 of equal catchability has been discussed with respect to closed population models, and we listed a number of examples of unequal catchability in bird populations. When identifiable subgroups of animals exhibit different probabilities of capture, then the groups can be treated separately. However, when variation in capture probability is present within a subgroup, problems can arise. Carothers (1973) examined the relative bias of the Jolly-Seber \(\hat{N}_i\) resulting from variation in catchability among individuals. When capture probability varied
moderately among individuals, the resulting relative bias of \( \hat{N}_i \) was fairly small for the situations examined, but when capture probability varied widely, large negative biases of \( \hat{N}_i \) resulted (Carothers 1973). Gilbert (1973) also used analytic approximations and computer simulation to examine the bias of \( \hat{N}_i \) associated with unequal catchability. Gilbert’s work emphasized the importance of the average capture probability. If this average is above 0.50, then bias resulting from variation among individuals will be small (Gilbert 1973). Gilbert (1973:524) concluded that “an experimenter need not attempt to design an experiment so that all animals have the same probability of capture, but only that the distribution be shifted nearer one so that nearly all animals have probabilities of capture say \( \geq 0.50 \)” A test of the equal catchability assumption in a K-sample experiment was provided by Leslie (1958) and extended by Carothers (1971) (also see the test of Cormack 1966). Practical recommendations for trying to avoid unequal catchability have been discussed relative to the two-sample experiment, although some of these suggestions (e.g., different capture techniques) may not be as applicable to the K-sample situation.

Assumptions 2 and 3 require a homogeneous population with regard to probability of surviving the intersample and sampling periods, respectively. Homogeneous sampling period survival probability can be assured by use of proper capture and handling techniques. However, the population can always be stratified if accidental deaths are thought to be higher among some groups of animals than others. Heterogeneity in intersample survival, \( \phi_i \), may be more difficult to account for or control. Age- and sex-specific variation in survival rates is thought to be common among birds (see Ricklefs 1973 and references therein). High predation rates on nesting females (e.g., see Johnson and Sargeant 1977) may also result in different survival probabilities of breeding and non-breeding adults. In any case, when survival probabilities differ among identifiable subgroups within a population, stratification is again appropriate. Certain handling and marking techniques are thought to influence survival probabilities of various bird species (see examples provided in the discussion of two-sample model assumptions). Tests of the hypothesis that survival is independent of capture probability and mark status were developed by Rob-son (1969), Manly (1971b), and Seber (1973:230–

**FIGURE 1.** Relationship between capture probability, \( P \), and the estimated coefficient of variation of \( \hat{N}_i \), \( CV(\hat{N}_i) \), assuming a six-period experiment with \( \phi_i = 0.50 \).
232). Robson (1969) discussed models that permit survival probabilities to be different for one or two sampling intervals after initial capture. These models were designed for field experiments in which marking or handling somehow stresses the captured animals. Pollock (1975a) generalized this model to permit not only different survival probabilities but also different capture probabilities for various numbers of intervals after initial capture. This very general model includes the Jolly-Seber model as a special case.

We have already noted that when subgroups (e.g., based on age, sex, breeding status) differ in capture or survival probability, they can be analyzed separately. However, when variation is age-specific young animals often have to be omitted from analysis until they become adults. Manly and Parr (1968) suggested one approach to estimating size of age-stratified populations. More recently, Pollock (1981b) and Stokes (1980) have developed general models for open, age-stratified populations. A model permitting geographic stratification, with animals of different areas having different capture and survival probabilities, has also been developed (Arnason 1973).

Assumptions 4 and 5 regarding retention of marks and reporting of recaptures have been discussed in the section on closed models. The assumption 6 of instantaneous sampling will of course never be strictly met, but efforts should be made to keep the sampling period fairly short relative to the intersample period. Mortality during the sampling period should be negligible. Assumption 7 regarding non-permanent emigration has not been well-studied but may be extremely important. Cormack (1981, pers. comm.) has had some success in recognizing nonpermanent emigration with his loglinear models approach. In addition, capture and/or resighting efforts in areas peripheral to the main study area could be used to gain insight to the magnitude of the problem. Detailed studies of movements of small samples of birds via radio telemetry could also provide an indication of the extent to which non-permanent emigration might be occurring.

In addition to individual tests of specific underlying assumptions, tests designed to assess the reasonableness of the Jolly-Seber model for a given data set are also available. Seber (1973:223–224) suggested a goodness-of-fit test for the Jolly-Seber model based on the expected
numbers of individuals having each possible capture history. We have found this test to be somewhat impractical because considerable cell pooling is generally required for the data sets we have examined and because large experiments require computation of a large number of expected cell values. Jolly (in prep.) has recently suggested a goodness-of-fit test based on the expected number of individuals released at time $i$ and next caught at times $i + 1, i + 2, \ldots, K$. This test also requires some cell pooling, but we have found it useful with actual data sets (see later examples). Another approach to assessing the reasonableness of the underlying model was suggested by Leslie et al. (1953) to estimate numbers of first captures and thus indirectly assess model fit. Estimated numbers of first captures agreed fairly well with actual values in most years, and it was concluded that the model provided a reasonable description of the data set. Agreement of Jolly-Seber estimates with “visual estimates” was poor, and the authors cautioned against the sole use of either method (Sulzbach and Cooke 1979). Kautz (1977) pointed out problems associated with the use of the Jolly-Seber model on Band-tailed Pigeon (Columba fasciata) data combined from a number of different trap sites. He recommended that parameter estimation should be restricted to data from a specific trapping site (or from sites in close proximity to each other). Kautz (1977) then used data from a Colorado banding site to estimate population size, and suggested that variation in these estimates might be indicative of variation in the area from which birds were drawn to the banded trapping station.

We are interested in the potential applicability of the Jolly-Seber model to avian mark-recapture studies, but this has been difficult to evaluate because of the paucity of published avian studies in which it has been used. Here we will attempt to provide some information on the potential utility of the method by presenting selected parameter estimates from Jolly-Seber analyses of a number of mark-recapture data sets for birds. The data sets were obtained from both published and unpublished sources and represent a variety of species, capture methods, sampling intervals, etc. Information on study locations and methodologies is presented in Table 1. Jolly’s (1965) estimates were computed for parameters of interest with all data sets. $X^2$ goodness-of-fit test statistics as suggested by Jolly (in prep.) were computed for all data sets in which adequate degrees of freedom remained after cell pooling. Some data sets were available only in Leslie Method B Table summary form (Leslie and Chitty 1951), and these sets were only subjected to the $X^2$ goodness-of-fit test. Data sets for which individual capture history data were available permitted estimation of the
number of first captures occurring in each sampling period as suggested by Leslie et al. (1953). For these data sets we recorded the proportion of the sampling periods (for which estimates of first captures could be computed) in which the estimated 95% confidence interval for first captures covered the actual known number.

Arithmetic mean estimates of capture probability, survival probability and population size, and the range of estimated coefficients of variation of population size are presented in Table 2. These summary statistics provide some indication of the relationship between these parameters and the precision of the resulting population size estimates. The two Canada Goose (Branta canadensis) data sets (sets 2 and 3) had the highest capture probabilities, and the resulting $\bar{C}(N_i)$ were low. The Pintail data set (4) exhibited the lowest estimated capture probability, but because of the large population size and resultant banded sample sizes, the $\bar{C}(N_i)$ were not the highest encountered. Similarly, the Manx Shearwater data set (1) had relatively low capture probabilities, but because of the large sample sizes the $\bar{C}(N_i)$ were also low. The data sets containing smaller sample sizes (e.g., fewer than 500 banded individuals) tended to yield a wide range of values for $\bar{C}(N_i)$, with low coefficients of variation associated with sampling periods having high capture probabilities, and vice versa, as expected.

The $\chi^2$ goodness-of-fit test statistics indicated significant ($P < 0.05$) rejection of the model in 3 of the 7 data sets for which they could be computed. The most obvious rejection was obtained for the Manx Shearwater data set (1), for which sampling was thought to be nonrandom (Orians 1958). The proportion of estimated confidence intervals covering the actual number of new captures was less than 0.95 for six of seven data sets, indicating possible violations of model assumptions.

A useful method of evaluating the utility or reasonableness of a parameter estimate is to compare it with an independent estimate of the same, or perhaps a similar, parameter. For example, the suggestion from the $\chi^2$ test that the male Canada Goose data set (2) from Old Hickory Lake, Tennessee, did not fit the Jolly-Seber model was disturbing because of the high capture probability and general quality of this data set. The female data set (3) from the same location provided a means of checking both sets of estimates. Male and female Canada Geese are generally thought to exhibit equivalent survival probabilities and the estimated mean Jolly-Seber survival rates for the 2 sexes did not differ significantly. If hatching sex ratio is approximately 50% males and if first year survival rates are approximately equal for the sexes (e.g., see summary data in Bellrose 1976), then male and female population sizes should be roughly equivalent. Annual population size estimates and associated 95% confidence intervals for adult males and females are plotted in Figure 3. There is good correspondence between the male and female estimates for all years. We have also plotted mid-winter aerial estimates of total population size for the Old Hickory Lake Canada Goose flock (from Cromer 1978:54). These mid-winter estimates apply to a different time of the year than the Jolly-Seber estimates, and they contain both sexes and all age classes, so correspondence between these and the sex-specific adult estimates is not expected to be close. Nevertheless, the similar patterns of population increase are reflected in the plots, and we believe that the comparisons presented in Figure 3 increase the credibility of the Jolly-Seber estimates for this population.

**Partially Open Populations**

Mark-recapture experiments on populations that are open to both gains and losses perhaps have the greatest potential applicability to studies on the dynamics of avian populations. However, if the population is partially rather than completely open, it is advantageous in the interest of parsimony to use one of the earlier models of Darroch (1959), which are shown by Jolly (1965) to be special cases of the Jolly-Seber model. A population that is closed to death and emigration but open to recruitment and immigration probably represents an extremely rare experimental situation. Thus, we have chosen to consider populations that experience only losses and no gains during the experimental period. A banding study of birds during a non-breeding period, or, if young recruits to the population can be distinguished from older residents, even the breeding season could be reasonably modeled with this partial closure assumption.

Assumptions generally listed for the "death but no recruitment" model are the same as those of the Jolly-Seber model but with the addition of (8) the population is closed to recruitment. Both Darroch (1959) and Jolly (1965) considered models based on the multinomial distribution and treated $d_i$, $p_i$, and (in Jolly's case) $v_i$ as unknown parameters to be estimated. Population size, $N_i$, at time $i$ is treated as an unknown random variable. The population size estimate under this model is:

$$N_i = \frac{R_i Z_i}{r_i} + n_i \quad (i = 1, 2, \ldots, K - 1)$$

where $R_i$ is the number of marked animals released after sample $i$, $Z_i$ is the number of ani-
<table>
<thead>
<tr>
<th>Data set</th>
<th>Species</th>
<th>Study location</th>
<th>Study period</th>
<th>Sampling time</th>
<th>Sampling period duration</th>
<th>Number sampling periods</th>
<th>Inter-sample period</th>
<th>Data source</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Manx Shearwater</td>
<td>Skokholm Island, Wales</td>
<td>1946–53</td>
<td>April–Sept.</td>
<td>6 mo</td>
<td>9</td>
<td>1 yr</td>
<td>Orians (1958: Table 3)</td>
</tr>
<tr>
<td>2</td>
<td>Canada Goose</td>
<td>Old Hickory Lake, TN</td>
<td>1967–77</td>
<td>20 June–14 July</td>
<td>3 days</td>
<td>11</td>
<td>1 yr</td>
<td>Cromer (1978: Table 6)</td>
</tr>
<tr>
<td>3</td>
<td>Canada Goose</td>
<td>Old Hickory Lake, TN</td>
<td>1967–77</td>
<td>20 June–14 July</td>
<td>3 days</td>
<td>11</td>
<td>1 yr</td>
<td>Cromer (1958: Table 7)</td>
</tr>
<tr>
<td>4</td>
<td>Pintail</td>
<td>Pel &amp; Kutawagan Marshes, Saskatchewan</td>
<td>1955–58</td>
<td>9 July–24 July</td>
<td>2 weeks</td>
<td>4</td>
<td>1 yr</td>
<td>Anderson &amp; Sterling (1974: Table 3)</td>
</tr>
<tr>
<td>5</td>
<td>American Woodcock</td>
<td>Canaan Valley, WV</td>
<td>1965–69</td>
<td>July–Sept.</td>
<td>3 mo</td>
<td>5</td>
<td>1 yr</td>
<td>Clark (1978: Table 13)</td>
</tr>
<tr>
<td>6</td>
<td>American Woodcock</td>
<td>Canaan Valley, WV</td>
<td>1965–69</td>
<td>July–Sept.</td>
<td>3 mo</td>
<td>5</td>
<td>1 yr</td>
<td>Clark (1978: Table 13)</td>
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<td>7</td>
<td>Band-tailed Pigeon</td>
<td>Longmont, CO</td>
<td>1970–75</td>
<td></td>
<td>2–3 mo</td>
<td>6</td>
<td>1 yr</td>
<td>Kautz (1977: Table 13)</td>
</tr>
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<td>8</td>
<td>Acadian Flycatcher</td>
<td>Patuxent Wildl. Res. Center, Laurel, MD</td>
<td>1959–72</td>
<td>Late May–early July</td>
<td>2 mo</td>
<td>14</td>
<td>1 yr</td>
<td>C. S. Robbins (unpubl. data)</td>
</tr>
<tr>
<td>9</td>
<td>Least Flycatcher</td>
<td>Hubbard Brook</td>
<td>1969–79</td>
<td>Late May–early July</td>
<td>2 mo</td>
<td>11</td>
<td>1 yr</td>
<td>R. T. Holmes (unpubl. data)</td>
</tr>
<tr>
<td>10</td>
<td>Barn Swallow (Hirundo rustica)</td>
<td>Patuxent Wildl. Res. Center, Laurel, MD</td>
<td>1948–54</td>
<td>Late May–early July</td>
<td>2 mo</td>
<td>7</td>
<td>1 yr</td>
<td>C. S. Robbins (unpubl. data)</td>
</tr>
<tr>
<td>12</td>
<td>Swainson's Thrush</td>
<td>Hubbard Brook</td>
<td>1969–79</td>
<td>Late May–early July</td>
<td>2 mo</td>
<td>11</td>
<td>1 yr</td>
<td>R. T. Holmes (unpubl. data)</td>
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<td>13</td>
<td>Red-eyed Vireo (Vireo olivaceus)</td>
<td>Hubbard Brook</td>
<td>1969–78</td>
<td>Late May–early July</td>
<td>2 mo</td>
<td>10</td>
<td>1 yr</td>
<td>R. T. Holmes (unpubl. data)</td>
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<tr>
<td>14</td>
<td>Red-eyed Vireo</td>
<td>Patuxent Wildl. Res. Center, Laurel, MD</td>
<td>1959–70</td>
<td>Late May–early July</td>
<td>2 mo</td>
<td>12</td>
<td>1 yr</td>
<td>C. S. Robbins (unpubl. data)</td>
</tr>
<tr>
<td>15</td>
<td>Yellow Warbler (Dendroica petechia)</td>
<td>Point Pelee, Ontario</td>
<td>1958–64</td>
<td>May–June</td>
<td>6–23 days</td>
<td>7</td>
<td>1 yr</td>
<td>Roberts (1971: Tables 1, 2)</td>
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<td>16</td>
<td>American Redstart</td>
<td>Hubbard Brook</td>
<td>1969–79</td>
<td>Late May–early July</td>
<td>2 mo</td>
<td>11</td>
<td>1 yr</td>
<td>R. T. Holmes (unpubl. data)</td>
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<td>17</td>
<td>Seaside Sparrow</td>
<td>Stone Harbor, NJ</td>
<td>1969–78</td>
<td>Late May–July</td>
<td>2–3 mo</td>
<td>10</td>
<td>1 yr</td>
<td>C. B. Worth (unpubl. data)</td>
</tr>
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<td>18</td>
<td>White-throated Sparrow</td>
<td>Columbus, OH</td>
<td>1946</td>
<td>14 Oct–4 Nov</td>
<td>1 day</td>
<td>20</td>
<td>1 day</td>
<td>Borror (1948: Fig. 1)</td>
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### Table 2
Summary Statistics of Jolly-Seber Analyses of Data Sets Described in Table 1*

<table>
<thead>
<tr>
<th>Data set</th>
<th>Species</th>
<th>Age*</th>
<th>Sex†</th>
<th>Number individuals banded</th>
<th>Number recaptures</th>
<th>(\hat{p})</th>
<th>(\hat{s})</th>
<th>(\hat{N})</th>
<th>(\hat{CV}(\hat{N}))</th>
<th>Range of (\hat{CV}(\hat{N}))</th>
<th>(\chi^2)</th>
<th>df</th>
<th>p</th>
<th>Proportional CI coverage*</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Manx Shearwater</td>
<td>A</td>
<td>M&amp;F</td>
<td>20,017</td>
<td>5175</td>
<td>0.18</td>
<td>0.68</td>
<td>16,046</td>
<td>0.05–0.08</td>
<td>105.87</td>
<td>21</td>
<td>0.00</td>
<td></td>
<td>—</td>
</tr>
<tr>
<td>2</td>
<td>Canada Goose</td>
<td>A</td>
<td>M</td>
<td>622</td>
<td>712</td>
<td>0.80</td>
<td>0.81</td>
<td>142</td>
<td>0.05–0.09</td>
<td>4.96</td>
<td>1</td>
<td>0.03</td>
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<tr>
<td>3</td>
<td>Canada Goose</td>
<td>A</td>
<td>F</td>
<td>611</td>
<td>764</td>
<td>0.76</td>
<td>0.84</td>
<td>147</td>
<td>0.04–0.11</td>
<td>5.95</td>
<td>3</td>
<td>0.11</td>
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<tr>
<td>4</td>
<td>Pintail</td>
<td>A</td>
<td>M</td>
<td>18,820</td>
<td>238</td>
<td>0.02</td>
<td>0.59</td>
<td>272,105</td>
<td>0.22–0.25</td>
<td>0.02</td>
<td>1</td>
<td>0.88</td>
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<tr>
<td>5</td>
<td>American Woodcock</td>
<td>A</td>
<td>M</td>
<td>127</td>
<td>17</td>
<td>0.23</td>
<td>0.73</td>
<td>190</td>
<td>0.67–1.45</td>
<td>—</td>
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</tr>
<tr>
<td>6</td>
<td>American Woodcock</td>
<td>A</td>
<td>F</td>
<td>201</td>
<td>23</td>
<td>0.43</td>
<td>0.34</td>
<td>197</td>
<td>0.03–0.89</td>
<td>—</td>
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<tr>
<td>7</td>
<td>Band-tailed Pigeon</td>
<td>A</td>
<td>M&amp;F</td>
<td>1186</td>
<td>217</td>
<td>0.19</td>
<td>0.65</td>
<td>1312</td>
<td>0.18–0.33</td>
<td>4.23</td>
<td>5</td>
<td>0.52</td>
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<tr>
<td>8</td>
<td>Acadian Flycatcher</td>
<td>A</td>
<td>M&amp;F</td>
<td>345</td>
<td>121</td>
<td>0.62</td>
<td>0.42</td>
<td>61</td>
<td>0.13–0.47</td>
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<tr>
<td>9</td>
<td>Least Flycatcher</td>
<td>A&amp;Y</td>
<td>M&amp;F</td>
<td>237</td>
<td>56</td>
<td>0.73</td>
<td>0.42</td>
<td>51</td>
<td>0.04–1.23</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td></td>
<td>0.38</td>
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<tr>
<td>10</td>
<td>Barn Swallow</td>
<td>A</td>
<td>M</td>
<td>134</td>
<td>74</td>
<td>0.63</td>
<td>0.75</td>
<td>61</td>
<td>0.08–0.73</td>
<td>—</td>
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<tr>
<td>11</td>
<td>Tufted Titmouse</td>
<td>A</td>
<td>M&amp;F</td>
<td>330</td>
<td>119</td>
<td>0.66</td>
<td>0.53</td>
<td>62</td>
<td>0.08–1.02</td>
<td>—</td>
<td>—</td>
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<tr>
<td>12</td>
<td>Swainson’s Thrush</td>
<td>A&amp;Y</td>
<td>M&amp;F</td>
<td>149</td>
<td>68</td>
<td>0.54</td>
<td>0.57</td>
<td>46</td>
<td>0.22–1.25</td>
<td>—</td>
<td>—</td>
<td>—</td>
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<td>—</td>
</tr>
<tr>
<td>13</td>
<td>Red-eyed Vireo</td>
<td>A&amp;Y</td>
<td>M&amp;F</td>
<td>201</td>
<td>95</td>
<td>0.42</td>
<td>0.57</td>
<td>77</td>
<td>0.14–0.67</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td></td>
<td>—</td>
</tr>
<tr>
<td>14</td>
<td>Red-eyed Vireo</td>
<td>A</td>
<td>M&amp;F</td>
<td>896</td>
<td>401</td>
<td>0.46</td>
<td>0.59</td>
<td>241</td>
<td>0.13–0.28</td>
<td>17.32</td>
<td>7</td>
<td>0.02</td>
<td>0.44</td>
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<tr>
<td>15</td>
<td>Yellow Warbler</td>
<td>A&amp;Y</td>
<td>M&amp;F</td>
<td>208</td>
<td>35</td>
<td>0.27</td>
<td>0.62</td>
<td>171</td>
<td>0.62–0.80</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.33</td>
<td>—</td>
</tr>
<tr>
<td>16</td>
<td>American Redstart</td>
<td>A&amp;Y</td>
<td>M&amp;F</td>
<td>232</td>
<td>55</td>
<td>0.37</td>
<td>0.67</td>
<td>66</td>
<td>0.08–1.13</td>
<td>—</td>
<td>—</td>
<td>—</td>
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</tr>
<tr>
<td>17</td>
<td>Seaside Sparrow</td>
<td>Y&amp;A</td>
<td>M&amp;F</td>
<td>230</td>
<td>112</td>
<td>0.60</td>
<td>0.55</td>
<td>60</td>
<td>0.14–0.42</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.83</td>
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<tr>
<td>18</td>
<td>White-throated Sparrow</td>
<td>A&amp;Y</td>
<td>M&amp;F</td>
<td>114</td>
<td>150</td>
<td>0.23</td>
<td>0.92</td>
<td>67</td>
<td>0.20–0.70</td>
<td>18.74</td>
<td>12</td>
<td>0.10</td>
<td>1.00</td>
<td>—</td>
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</table>

* \(\hat{p}\) denotes arithmetic mean capture probability estimate. \(\hat{s}\) denotes arithmetic mean survival probability estimate. \(\hat{N}\) denotes arithmetic mean population size estimate, and \(\hat{CV}(\hat{N})\) denotes the estimated coefficient of variation for population size in sampling period \(i\).

* Y denotes young (first year) and A denotes adult.

† M denotes male and F denotes female.

* Test suggested by Jolly (in prep.)

[\(\hat{N}_i = \frac{1}{2}(\hat{N}_{i-1} + \hat{N}_i)\).]
mals not caught in sample \( i \) but caught subsequently, \( r_i \) denotes the number of marked animals released after sample \( i \) and subsequently recaptured, and \( n_i \) is the number of animals caught in sample \( i \). This \( N_i \) can be shown to be an intuitively reasonable estimator using logic analogous to that used previously for the Jolly-Seber estimator, \( M_i \).

This model has not been widely used by ecologists. Robson (1979) has discussed the planning of experiments designed to estimate survival rates using this model, but we are aware of no similar work emphasizing estimation of \( N_i \). We do note that examination of the estimator for the variance of \( N_i \) (see Jolly 1965:242) shows the importance of a large number of recaptures, \( r_i \), to the precision of \( N_i \). For this reason, the suggestion of Jolly (1965:239) for his completely open model that release and recapture operations can be operated independently is relevant to this model also. While estimates of \( N_i \) can only be obtained for sampling periods in which animals are released, recaptures could be obtained continuously (e.g., via resightings) by biologists or even volunteers in order to improve the precision of \( N_i \).

The discussion of assumptions 1–7 for completely open models is relevant to the "death but no immigration" model also. Pollock et al. (1974) provide a series of tests which can be used to examine assumption 8 of partial closure. Proper choice of time of the year and duration of the experiment can serve to improve the chance that this assumption is met. Finally, we note that a \( \chi^2 \) goodness-of-fit test for the model can be computed in a manner analogous to that suggested by Jolly (in prep.) for the general Jolly-Seber model.

To our knowledge, the only use of this model in ornithological work has been that of Dwyer (pers. comm.). He has used the model to estimate population size of Woodcock based on summer mark-recapture experiments. The model was thought to produce reasonable estimates, and goodness-of-fit test statistics indicated that most of the data sets did fit the model.

**DISCUSSION**

As indicated in this review, ornithologists have not made much use of mark-recapture methodologies for estimating population size. A modest number of reviewed studies employed the two-sample Lincoln index experiment which unfortunately does not permit much of the testing of assumptions which is possible with \( K \)-sample experiments. Ornithologists have made virtually no use of the models developed for \( K \)-sample experiments on closed populations. A small number of examples of the use of \( K \)-sam-

![Figure 3](image-url)
instructions, output explanation, and examples of the use of two user-oriented FORTRAN programs for carrying out necessary computations.

A number of recent efforts have been directed at making mark-recapture methods both understandable and accessible to biologists. Seber's (1973) book (a new edition is due in 1980) provides an excellent review of methodology. Methods are illustrated with numerous field examples and much of the book should be understandable to biologists. More recently, Begon (1979) has written a monograph on capture-recapture methods aimed specifically for biologists. Two papers by Cormack (1972, 1973) are directed at the logic of mark-recapture methods and the intuitive nature of the associated estimators. The monograph of Otis et al. (1978) on closed population models is also written with the biologist in mind. A more elementary treatment of the material presented in Otis et al. (1978) is provided by White et al. (1981).

With respect to computational algorithms, computer programs providing Jolly-Seber estimates have been available for nearly a decade (Davies 1971, White 1971, Arnason and Kreeger 1973). The new version (see Arnason and Banjuk 1980) of one of these programs has extensive data management capabilities and also computes estimates based on the two partially open population models of Jolly (1965) and the closed population model of Darroch (1958). A comprehensive computer program was developed by Otis et al. (1978) to provide estimates and conduct tests leading to selection of appropriate closed population models (see White et al. 1978). Crosbie (1979) has developed a computer program which computes estimates based on a number of open population models (e.g., the standard Jolly-Seber model and similar reduced-parameter models) and computes test statistics to aid in the selection of an appropriate model. We have developed a program for the age-stratified open population model of Stokes (1980) which is available to interested researchers now and which should be sufficiently user-oriented for general distribution in the near future (Hines, Stokes and Nichols, unpubl.).

It is difficult to make general statements about the potential applicability of mark-recapture methods to avian population estimation because of the small number of relevant studies which have been conducted. In particular, there has been a complete lack of K-sample closed population experiments. We hope that the models and program of Otis et al. (1978) will be used by ornithologists for short-term experiments (or in conjunction with long-term investigations comprised of a number of relatively short sampling periods), and we will be interested to learn how well the various models seem to fit bird recapture data. With respect to open populations, the analyses presented in Table 2 provide some indication of the applicability of the Jolly-Seber model to avian studies. Results from some of the data sets (e.g., the Canada Geese) were very encouraging. However, assessments of model fit provided indications of problems with underlying assumptions in a number of the data sets. In addition, precision of population size estimates varied widely within a number of the data sets. We suspect that the age-stratified open population models of Pollock (1981b) and Stokes (1980) will be useful in avian studies, and it will be interesting to learn whether the general models of Robson (1969) and Pollock (1975a) are useful with bird recapture data. The large variance estimates associated with many of our Jolly-Seber analyses emphasize the need to consider the parsimonious, reduced-parameter models of Jolly (1979, in prep.) and Crosbie (1979). We hope that ornithologists interested in long-term population studies will examine these various open population models and make use of them as they become available.

ACKNOWLEDGMENTS

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